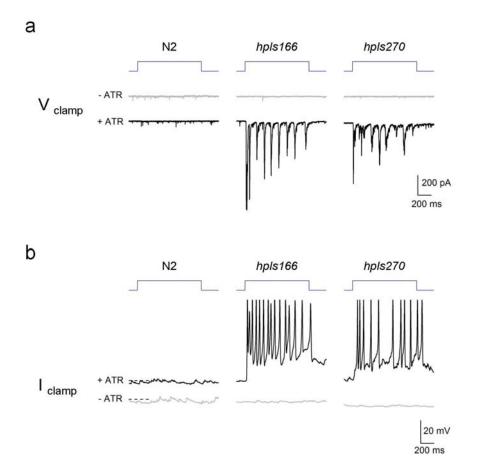


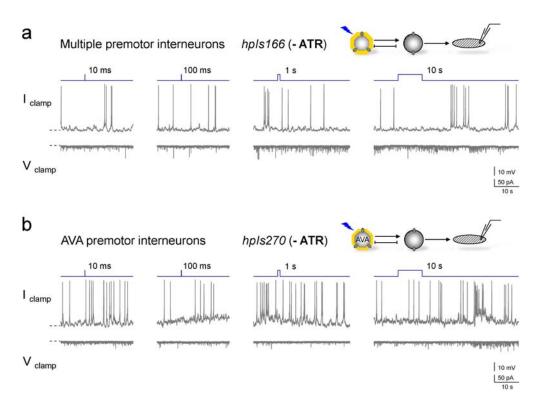
$Supplementary\ Figure\ 1.\ Difference\ between\ miniature\ PSCs\ versus\ rPSCs\ events.$

(a) Example traces of spontaneous miniature PSCs (minis, left), and rhythmic bursting PSCs (rPSCs, right) in wild-type animals. Body wall muscles were held at -60 mV. The enlarged view of each trace is shown below. (b,c) Sample traces of monophasic and multiphasic miniature PSCs events. (d) Sample traces of single rPSCs events.



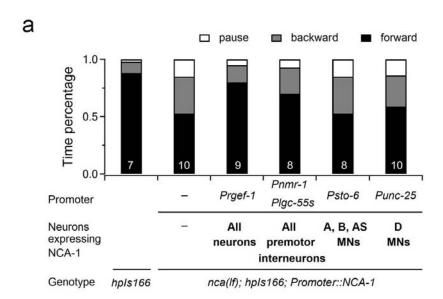
Supplementary Figure 2. rPSCs correlate with action potentials at body wall muscles.

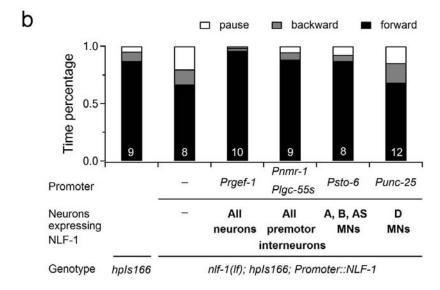
(a) Representative rPSCs traces, from animals of various genetic backgrounds (N2, *hpls166* and *hpls270*), cultured with or without *all-trans* retinal (+ATR, -ATR, respectively), upon 1 second blue light stimulation. Body wall muscles were held at -60 mV. (b) Representative action potential traces recorded from these animals. Body wall muscles were held at 0 pA. Dash lines denote -25 mV.



Supplementary Figure 3. No persistent activity was observed in the absence of ATR.

(**a,b**) Recording of the same transgenic strains (**a** *hpIs166*, **b** *hpIs270*) and by the same stimulation protocols, as in Figure 1**c**, cultured in the absence of *all-trans* retinal (-ATR). V $_{clamp}$: 10 ms (n = 7), 100 ms (n = 7), 1 s (n = 10), 10 s (n = 7); I $_{clamp}$: 10 ms (n = 64), 100 ms (n = 6), 1 s (n = 6), 10 s (n = 5) (**a**, quantified in Figure 1**d**); V $_{clamp}$: 10 ms (n = 6), 100 ms (n = 6), 1 s (n = 8), 10 s (n = 9); I $_{clamp}$: 10 ms (n = 7), 100 ms (n = 7), 1 s (n = 7), 10 s (n = 7) (**b**, quantified in Figure 1**f**).





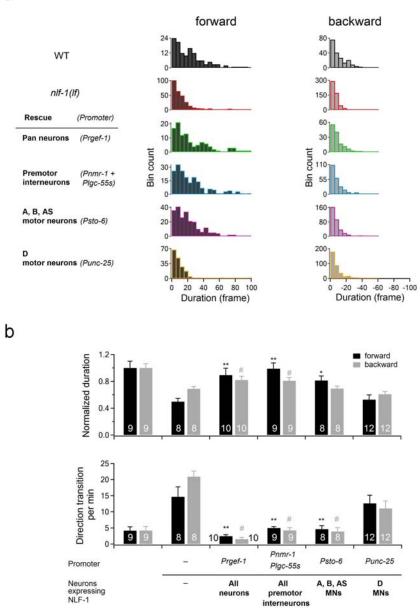
Supplementary Figure 4. A key requirement of NCA in premotor interneurons.

(a) Under the same assay conditions (Methods), the spontaneous motor activity was reduced in nca(lf) mutants, represented by the increased percentage of time animals spent in the pausing (white bars, centroid moved between -1 and 1 pixel/second) mode. The reduced motor activity was rescued by restoring the NCA channel expression in premotor interneurons, but not in motor neurons. (b) Similar defects were observed in nlf-1 mutant animals. Restored expression of NLF-1 in either premotor interneurons or excitatory motor neurons could rescue the overall activity level. All strains were analyzed in the hpIs166 transgenic background.

Genotype

hpls166

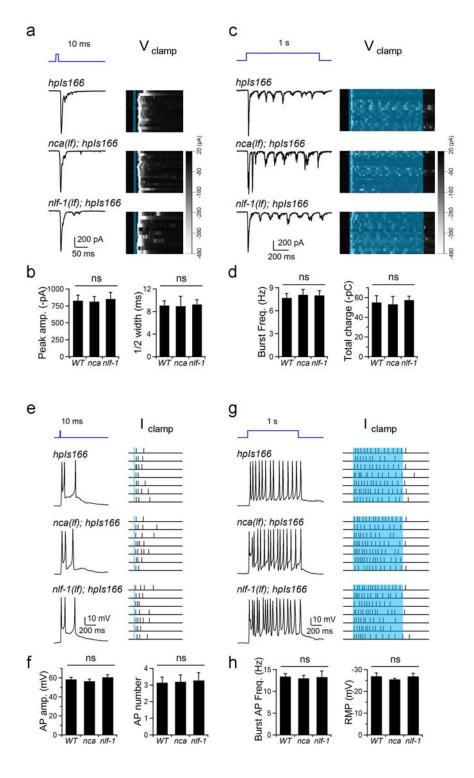
a



nlf-1(lf); hpls166; Promoter::NLF-1

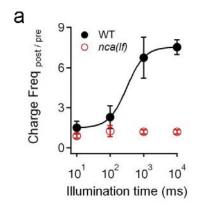
Supplementary Figure 5. NLF-1 is required for sustained locomotion.

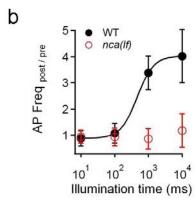
(a) Histogram of the duration of sustained directional movement (forward and backward) in animals of different genetic backgrounds. The duration of sustained forward and backward movements was significantly reduced in nlf-1(lf) mutant animals. This defect was rescued by restoring expression of NLF-1 in premotor interneurons (Pnmr-1 + Plgc-55s). (b) Quantification of the average duration and frequency of directional transition. NLF-1 expression in premotor interneurons was necessary for increasing the duration (both forward and backward) for nlf-1(lf) mutants. Restoring expression of NLF-1 in cholinergic motor neurons (Psto-6) can partially rescues the forward duration, but not backward duration, whereas expression of NLF-1 in GABAergic motor neurons (Punc-25) did not rescue the duration of forward or backward movement. All strains were analyzed in the hpIs166 transgenic background. * P < 0.05, ** P < 0.01, # P < 0.05, against nlf-1(lf) mutants by the Mann-Whitney U test. Error bars, SEM.



Supplementary Figure 6. Similar evoked activity in wild-type, *nca(lf)* and *nlf-1(lf)* animals.

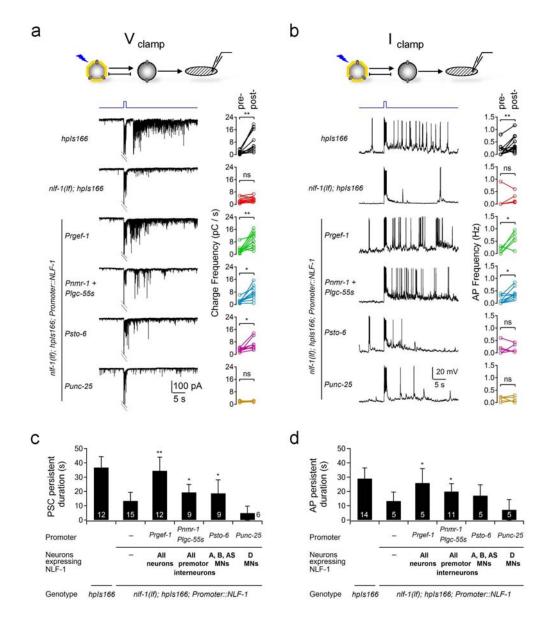
(a) Left panel, representative traces of rPSCs evoked during 10ms-blue light stimulation in wild-type (WT), nca(lf) and nlf-1(lf) animals, respectively. Right panel, raster plots of rPSCs evoked by 10ms-blue light stimulation from each sample. (b) Quantifications of the peak amplitude and half-width of rPSCs. There are no significant difference between WT animals and nca(lf) and nlf-1(lf) mutants. WT (n = 15), nca(lf) (n = 17), nlf-1(lf) (n = 11). (c) Left panel, representative traces of rPSCs evoked during 1s-blue light stimulation in wild-type, nca(lf) and *nlf-1(lf)* animals, respectively. 1s-blue light stimulation induced rPSCs in all three genotypes. Right panel, raster plot of individual recordings. (d) Quantifications of the frequency and total discharge of rPSCs. There are no significant difference between wild-type animals, nca(lf) and nlf-1(lf) mutants. WT (n = 16), nca(lf) (n = 19), nlf-1(lf) (n = 15). (e-h) Example traces and quantification of APs evoked by 10ms- and 1s- blue light stimulation, in I clamp configuration. There was no significant difference among these genotypes. WT (n = 12), nca(lf) (n = 7), nlf-I(lf) (n = 9) (**f**). WT (n = 18), nca(lf) (n = 9), nlf-I(lf) (n = 7) (**h**). All recordings were performed in the *hpIs166* background (wild-type animals in this set of experiments). Muscles were held at -60 mV in V clamp and 0 pA in I clamp, respectively. ns, no significant difference against wild-type animals by the Mann-Whitney U test. Error bars, SEM.





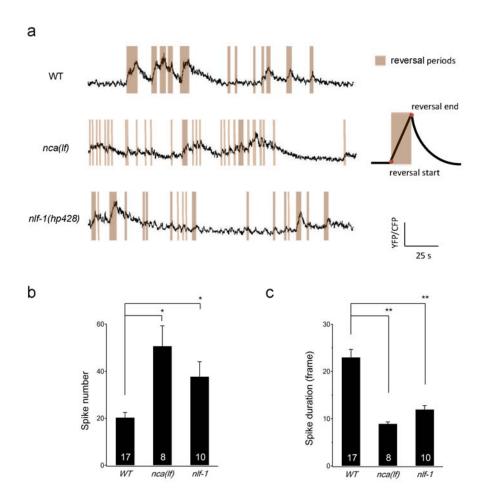
Supplementary Figure 7. nca(lf) mutants exhibit no persistent neural activity.

nca(lf) mutants did not exhibit increased charge frequency (rPSCs; **a**) or firing frequency (action potentials; **b**), with illumination times from 10ms to 10s. V $_{clamp}$: 10 ms (WT, n = 10; nca(lf), n = 13), 100 ms (WT, n = 7; nca(lf), n = 11), 1 s (WT, n = 12; nca(lf), n = 16), 10 s (WT, n = 10; nca(lf), n = 21) (**a**); I $_{clamp}$: 10 ms (WT, n = 14; nca(lf), n = 7), 100 ms (WT, n = 9; nca(lf), n = 7), 1 s (WT, n = 14; nca(lf), n = 7), 10 s (WT, n = 7; nca(lf), n = 4) (**b**). Error bars, SEM. Data were collected from more than four biological and technical replicates (independent culture preparations). Error bars, SEM.



Supplementary Figure 8. NLF-1 is required for persistent activity.

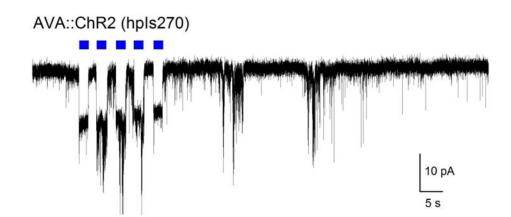
(a) Left panels, representative traces of persistent rPSCs after 1s-blue light stimulation. Right panels, rPSCs charge frequency before and after the stimulation. Connected lines represent the recording from same preparation. (b) Quantification of the duration of persistent rPSCs. Restoring expression of NLF-1 in premotor interneurons provided the best restoration of persistent neural activity in nlf-1 mutants. Restoring expression of NLF-1 in all cholinergic motor neurons (Psto-6) partially rescued the deficit, whereas restored NLF-1 expression in GABAergic motor neurons (Punc-25) did not rescue the defects. (c,d) Representative traces and quantification of AP frequency before and after the stimulation. The schematics of the recording configuration (V_{clamp} (a) and I_{clamp} (c)) were shown. All recordings were performed in hpIs166 background (wild-type animals in this set of experiments). Muscles were held at -60 mV in V_{clamp} and at 0 pA in I_{clamp} , respectively. * P < 0.05, ** P < 0.01, ns, no significant difference against nlf-1(lf) mutants by the Mann-Whitney U test. Error bars, SEM.



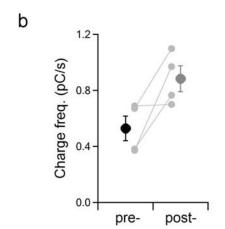
Supplementary Figure 9. Fainter mutants exhibit reduced Ca²⁺ transient duration.

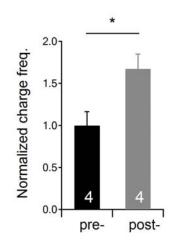
(a) Ca^{2+} transients in coimaged AVA/AVE indicated by the YFP/CFP ratio of a cameleon reporter in wild type and fainter animals. Brown bars indicate backing locomotion. (b) Quantification of the number of Ca^{2+} spikes. Fainter mutants exhibit high frequency initiation that correlates with increased number of Ca^{2+} spikes. (c) Quantification of the duration of Ca^{2+} increase. Fainter mutants exhibit reduced Ca^{2+} spike width. * P < 0.05, ** P < 0.01 by the Mann-Whitney U test. Error bars, SEM

а



С





Supplementary Figure 10. AVA exhibits persistent activity upon stimulation.

(a) AVA recording in a dissected *hpIs270* preparation (hold at -60 mV by whole cell patch clamping) upon a train of 2s 470nm light stimulation at 2s interval. Solutions used for AVA recording are the same as those used for the neuromuscular preparation. (b) Charge frequency of AVA was significantly increased after stimulation in 3 of 4 such preparations. Charge transfer during 10 second pre- to 40-60 second post-stimulation was quantified. Connected lines represent recordings from the same neuron before and after the stimulation. (c) Normalized charge frequency showed significantly difference before and after the stimulation. * P < 0.05 by Student's t test. Error bars, SEM.

Supplementary Table 1. Correlation between persistent activity and locomotion.

